

**Review Article****Multifaceted Roles of *Trichoderma* in Sustainable Agriculture: Mechanisms, Applications, and Future Perspectives**Sujata Nepal^{1*}, Surakshya Sharma², Niraj Mahato³¹Institute of Agriculture and Animal Science, Rampur Campus, Khairahani Chitwan, Nepal.²Institute of Agriculture and Animal Science, Kirtipur, Kathmandu, Nepal.³Agriculture and Forestry University, central campus, Rampur, Chitwan, Nepal.

*Correspondence: (Sujata Nepal)

sujatanepal58@gmail.com**Abstract**

Modern agricultural practices have boosted crop yields but have also intensified pressure on the food system, along with environmental and health issues linked to overreliance on chemical fertilizers and pesticides. Soil degradation, loss of biodiversity, pesticide resistance, pollution, and human health hazards are the serious negative consequences imposed due to intensive agricultural practices, necessitating the shift towards biological agents to boost productivity and safeguard environmental and human health. Beneficial organisms, especially *Trichoderma* species, have emerged as effective beneficial fungi due to their versatile roles in sustainable agriculture for disease suppression through mycoparasitism, competition, production of secondary metabolites, and entomopathogenesis. In addition to pathogen suppression, *Trichoderma* spp. induce defence mechanisms in plants, produce growth hormones, mobilize unavailable nutrients, and increase nutrient uptake, making plants tolerant to biotic and abiotic stress and facilitating the bioremediation of toxic soil. However, problems related to strain specificity, field performance, environmental conditions and shelf-life stability limit its widespread adoption. Future studies should focus on producing stress-tolerant and highly efficient strains, *Trichoderma* strains that can tolerate broader environmental conditions, exploring synergetic effects with other beneficial micro-organisms, and application methods. Overall, this review presents the versatile function of *Trichoderma* spp. in increasing crop yield and preventing negative consequences on environmental and human health, and also highlights challenges and the need for advance future studies.

Keywords: Biocontrol, Biofertilizer, Plant Growth Promotion, Soil Microbiome Interactions, Sustainable Agriculture

1. Introduction

Modern agriculture has become increasingly dependent on intensive farming practices involving the excessive application of chemical fertilizers and pesticides to maximize crop productivity, and fulfill the growing food demand [1–3]. The rapid response, easy availability, and immediate effectiveness of these agrochemicals have contributed significantly to their widespread adoption among farmers [4]. Although these practices have substantially enhanced agricultural output, their prolonged and indiscriminate use has generated severe environmental, ecological, and human health concerns.

The continuous application of chemical fertilizers and pesticides has adversely affected soil fertility, biodiversity, and environmental quality. Excessive pesticide application contaminates soil and water bodies, leading to eutrophication, destruction of beneficial soil microflora, and mortality of non-target organisms [5, 6]. Similarly, the overuse of synthetic fertilizers contributes to the accumulation of toxic compounds and heavy metals in agricultural soils, negatively affecting soil health, crop quality, and food safety [7]. Furthermore, prolonged use of pesticides with similar modes of action has accelerated the development of resistant strains of pathogens

and insect pests, reducing the effectiveness of chemical control measures [8]. These environmental and agronomic challenges highlight the urgent need for sustainable agricultural practices capable of maintaining crop productivity while minimizing ecological degradation.

In recent years, sustainable and eco-friendly agricultural approaches have gained considerable attention as alternatives to chemical-intensive farming systems. Among these approaches, the use of beneficial microorganisms as biofertilizers and biocontrol agents has emerged as a promising strategy for enhancing crop productivity and maintaining environmental sustainability [9, 10]. Unlike chemical pesticides, microbial biocontrol agents generally possess multiple mechanisms of action, exhibit target specificity, and pose minimal risk to human health and the environment [3, 11]. These characteristics reduce the likelihood of resistance development among plant pathogens and pests, making microbial-based technologies a sustainable option for long-term agricultural management.

Among the diverse groups of beneficial microorganisms, *Trichoderma* spp. have attracted significant scientific and commercial interest due to their multifunctional roles in

sustainable agriculture. Species of *Trichoderma* are widely distributed soil fungi known for their rapid growth, high reproductive capacity, adaptability to diverse environmental conditions, and strong rhizosphere colonization ability [12]. They can exist as saprophytes in soil organic matter, endophytes within plant tissues, and opportunistic symbionts associated with plant roots [13]. Upon colonization, *Trichoderma* spp. produce a wide range of bioactive compounds, hydrolytic enzymes, and secondary metabolites that promote plant growth, suppress pathogens, improve nutrient uptake, and induce plant defense responses [14–16].

In addition to their role as biocontrol agents against fungal pathogens, nematodes, and insect pests, *Trichoderma* spp. contribute to nutrient solubilization, phytohormone production, abiotic stress tolerance, and bioremediation of contaminated soils [17, 18]. Their broad-spectrum activities make them valuable components of integrated and sustainable agricultural systems aimed at reducing dependency on synthetic agrochemicals.

Therefore, this review aims to provide a comprehensive overview of the multifaceted roles of *Trichoderma* spp. in sustainable agriculture. The review focuses on their mechanisms of action, applications as biocontrol agents and biofertilizers, roles in plant growth promotion and stress tolerance, bioremediation potential, and the environmental factors influencing their effectiveness. Additionally, the review highlights current challenges and future prospects for the successful integration of *Trichoderma*-based technologies into sustainable crop production systems.

2. Taxonomy, Characteristics, and Ecology of *Trichoderma*

2.1. Taxonomy and Classification

Trichoderma is a filamentous fungal genus belonging to the kingdom Fungi, phylum Ascomycota, class Sordariomycetes, order Hypocreales, and family Hypocreaceae [19]. Species of *Trichoderma* are commonly associated with the sexual state Hypocrea, although many agriculturally important strains are primarily recognized and utilized in their asexual forms [20]. Traditionally, *Trichoderma* species were categorized under fungi imperfecti because of the absence or rarity of sexual reproduction; however, advances in molecular taxonomy and phylogenetic analyses have significantly improved species identification and classification [21, 22].

Trichoderma includes more than 500 species, and ongoing taxonomic studies continue to add new species [23]. Among them, *T. harzianum*, *T. virens*, *T. atroviride*, *T. asperellum*, *T. asperelloides*, *T. hamatum*, *T. longibrachiatum*, and *T. gamsii* are the most extensively studied species because of their strong biocontrol potential and plant growth-promoting properties [24–28]. Due to their multifunctional capabilities and adaptability, these species are widely utilized in agriculture as biological control agents, biofertilizers, and bio-stimulants [29].

2.2. Morphological and Physiological Characteristics

Species of *Trichoderma* are characterized by rapid growth, profuse sporulation, and efficient colonization ability. They produce fast-growing, branched, septate hyphae with green-colored conidia that serve as a distinguishing morphological feature of the genus [30, 31]. Colonies are generally green to dark green due to abundant conidial production and can grow on a wide range of nutrient media, including potato dextrose agar [PDA], malt agar [MA], and Czapek-Dox agar [CDA] [20, 32].

Trichoderma spp. are highly adaptable fungi capable of surviving under diverse environmental conditions. Most species exhibit optimal growth between 25–30°C and prefer slightly acidic conditions, although some strains can tolerate broader temperature and pH ranges [33, 34]. Their remarkable adaptability and rapid reproductive capacity contribute to their ecological success and effectiveness in agricultural systems. Physiologically, *Trichoderma* spp. possess strong enzymatic machinery and metabolic versatility. They produce several extracellular enzymes, including chitinases, glucanases, cellulases, and proteases, which facilitate decomposition of organic matter and antagonism against plant pathogens [29, 35]. In addition, these fungi synthesize a wide range of secondary metabolites, volatile organic compounds, phytohormones, and siderophores that contribute to plant growth promotion, nutrient mobilization, and disease suppression [9, 36].

2.3. Ecology and Rhizosphere Colonization

Trichoderma species are cosmopolitan fungi commonly found in agricultural soils, forest ecosystems, decaying wood, compost, and other organic substrates [1]. Due to their saprophytic nature and rapid rhizosphere colonizing ability, *Trichoderma* create a strong competition for space and mobilizing nutrient from soil in comparison to other soil organisms [29, 35]. Besides their saprophytic nature, many species can establish endophytic associations with plants by colonizing root tissues without causing disease symptoms [13]. One of the most important ecological characteristics of *Trichoderma* spp. is their exceptional rhizosphere competence. They effectively colonize plant root surfaces and surrounding soil through rapid spore germination, hyphal proliferation, and competition for nutrients and space [29]. During root colonization, *Trichoderma* interacts closely with plants through chemical signaling and secretion of bioactive compounds that influence plant physiology and defense mechanisms [15].

The successful establishment of *Trichoderma* in the rhizosphere enhances nutrient availability, stimulates root development, and improves plant resistance against biotic and abiotic stresses [16]. Moreover, their ability to survive under fluctuating environmental conditions and interact synergistically with other beneficial microorganisms makes them important components of sustainable agricultural ecosystems [37].

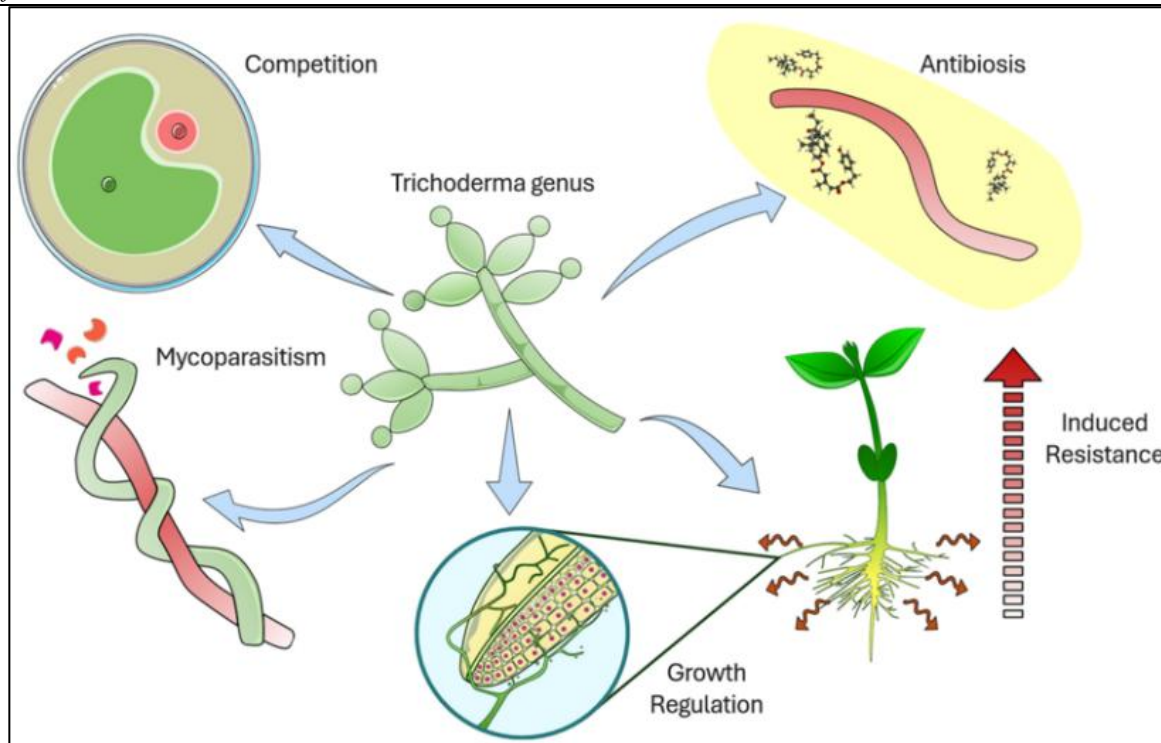


Figure 1. Mechanism of action for biocontrol exhibited by *Trichoderma* spp. Reproduced from [38].

3. Mechanisms of Action of Trichoderma

The effectiveness of *Trichoderma* spp. in sustainable agriculture is primarily attributed to their diverse and synergistic mechanisms of action against plant pathogens and environmental stresses. These mechanisms involve both direct and indirect interactions with pathogens, insects, nematodes, and host plants. Depending on the species, strain, host plant, and environmental conditions, *Trichoderma* may exhibit antagonistic activities through competition, mycoparasitism, antibiosis, induction of plant defense responses, and entomopathogenic or nematocidal actions [39–41]. The multifunctional nature of these mechanisms contributes significantly to the broad-spectrum efficacy of *Trichoderma* in crop protection and plant growth promotion. The mechanism of action for biocontrol exhibited by *Trichoderma* spp. is illustrated in figure 1.

3.1. Competition for Nutrients and Space

One of the primary mechanisms by which *Trichoderma* suppresses plant pathogens is through competition for nutrients and ecological niches in the rhizosphere. Due to their rapid growth rate, high reproductive capacity, and strong rhizosphere colonization ability, *Trichoderma* spp. effectively occupy root surfaces and surrounding soil regions, thereby limiting the availability of space and nutrients for pathogenic microorganisms [29, 35]. Iron is one of the most critical micronutrients involved in microbial competition within the soil environment [42]. *Trichoderma* spp. produce

siderophores, which are low molecular weight iron-chelating compounds that efficiently mobilize iron from the surrounding environment [43]. These siderophores deprive competing pathogens of available iron, restricting their growth and [36, 44]. In addition, rapid nutrient uptake and efficient carbon utilization further enhance the competitive advantage of *Trichoderma* over pathogenic fungi [45].

3.2. Mycoparasitism

Mycoparasitism is considered one of the most important direct antagonistic mechanisms of *Trichoderma* against phytopathogenic fungi. In this process, *Trichoderma* recognizes, attacks, and parasitizes the hyphae or resting structures of pathogenic fungi [46]. The initiation of mycoparasitism involves chemotropic growth toward the host fungus, followed by attachment, coiling around the pathogen hyphae, and penetration of the cell wall [35, 47].

The penetration process is facilitated by the secretion of extracellular hydrolytic enzymes such as chitinases, β -1,3-glucanases, cellulases, and proteases, which degrade the structural components of the pathogen cell wall [39]. These enzymatic activities ultimately result in lysis and destruction of the target pathogen. Studies have demonstrated that *T. harzianum* exhibits strong mycoparasitic activity against several economically important pathogens, including *Alternaria porri*, *Fusarium oxysporum*, *Rhizoctonia solani*, and *Sclerotium rolfsii* [30, 35]. Representative examples of plant diseases controlled by *Trichoderma* spp. are presented in table 1.

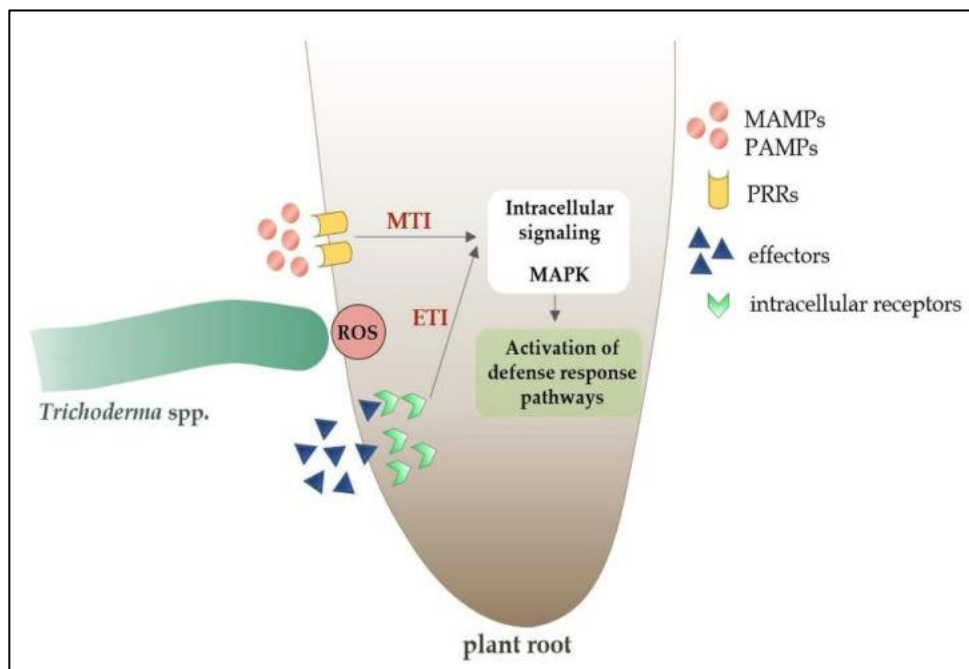


Figure 2. Plant defense response pathways triggered by microbe-associated molecular patterns (MAMPs) and effector molecules from *Trichoderma* spp. Reproduced from [48]. These pathways are consistent with recent reports demonstrating that *Trichoderma* spp. induce plant defense responses through complex mechanisms involving signaling molecules and pathogen suppression [49].

Table 1. Some examples of plant diseases controlled with *Trichoderma*.

Pathogen Species	Trichoderma Species	Mode of Action	References
<i>Botrytis cinerea</i>	<i>Trichoderma</i> spp.	Development of secondary metabolites	[50]
<i>Phytophthora capsici</i>	<i>T. virens</i>	Mycoparasitism and production of gliotoxin, a secondary metabolite.	[24]
<i>Colletotrichum gloeosporioides</i>	<i>T. asperelloides</i> <i>T. asperellum</i>	Competition and production of secondary metabolites	[25]
<i>Meloidogyne incognita</i> <i>Fusarium oxysporum</i>	<i>T. longibrachiatum</i>	Synthesis of Zinc nanoparticles, altering the expression of defense gene and destruction of hyphal growth.	[26]
<i>Diplodia seriata</i>	<i>T. harzianum</i> <i>T. carraovejensis</i> <i>T. gamsii</i>	Antibiosis through the production of bio active secondary metabolites and extra cellular enzymes.	[3]
<i>Pyricularia oryzae</i>	<i>T. harzianum</i>	Not mentioned	[4]
<i>Phytophthora cinnamomi</i>	<i>T. atroviride</i>	Competition	[51]
<i>Fusarium avenaceum</i>	<i>T. atroviride</i>	Growth inhibition of pathogen through the secretion of Volatile Organic Compound.	[52]
<i>Fusarium oxysporum</i>	<i>T. harzianum</i>	<i>Mycoparasitism</i>	[53]

3.3. Antibiosis

Antibiosis refers to the production of inhibitory compounds by one microorganism that adversely affect the growth or metabolism of another microorganism [54]. *Trichoderma* spp. are well known for producing a wide range of primary and

secondary metabolites with strong antifungal, antibacterial, and insecticidal activities. These includes 390 metabolites such as such as gliotoxin, gliovirin, heptelidic acid, and viridin. Sepedonin, catechin, caffeic acid, ferulic acid, 3, 4, 15-scirpenetriol and naematolin inhibits the growth and development of other fungal pathogens [9, 26, 55].

Table 2. Some examples of insects control by *Trichoderma*.

Pest Species	<i>Trichoderma</i> Species	Mode of Action	References
<i>Macrosiphum euphorbiae</i>	<i>T. atroviride</i>	Activation of Salicylic acid pathway which resulted on poor sap quality and attraction of parasitoid.	[56]
<i>Spodoptera littoralis</i>	<i>T. atroviride</i>	Activation of jasmonic acid pathway which resulted on inhibition of digestion and suppression of moth growth	[56]
<i>Aphis gossypii</i>	<i>T. harzianum</i>	Not mentioned	[57]
<i>Brevicoryne brassicae</i>	<i>T. harzianum</i>	Not mentioned	[27]
<i>Ceratovacuna lanigera</i> Zehntner	<i>T. hamatum</i>	Not mentioned	[58]
<i>Aedes aegypti</i>	<i>T. atroviride</i>	Direct infection through cuticle followed by production of secondary metabolites	[59]
<i>Spodoptera littoralis</i>	<i>T. hamatum</i>	Direct infection through natural openings	[60]
<i>Locusta migratoria</i>	<i>Trichoderma</i> spp.	Development of secondary metabolites	[50]
<i>Diuraphis noxia</i>	<i>T. harzianum</i>	Development of secondary metabolites	[28]
<i>Tribolium castaneum</i>	<i>T. harzianum</i>	Development of secondary metabolites	[28]
<i>Xylosandrus germanus</i>	<i>T. harzianum</i> , <i>T. asperellum</i> , and <i>T. atroviride</i>	Mycoparasitism on the symbiotic fungus <i>Ambrosiella grosmanii</i>	[61]

The antimicrobial compounds produced by *Trichoderma* interfere with cell membrane integrity, enzymatic activity, and metabolic pathways of pathogens, thereby inhibiting spore germination and mycelial growth [41]. Volatile organic compounds produced by certain strains of *Trichoderma* can also suppress pathogen growth without direct physical contact, thereby enhancing their effectiveness in the rhizosphere environment [62].

3.4. Induction of Plant Defense Responses

In addition to direct antagonistic effects on pathogens, *Trichoderma* spp. can enhance plant resistance by activating various defense mechanisms in host plants. Colonization of plant roots by *Trichoderma* triggers physiological and biochemical changes that stimulate both local and systemic defense responses [18, 63]

Several elicitors produced by *Trichoderma*, including hydrophobins, cellulases, glycoproteins, terpenoids, flavonoids, and polypeptides, function as signaling molecules that activate plant immune responses [64]. These elicitors stimulate the production of reactive oxygen species (ROS), phytoalexins, pathogenesis-related proteins, and defense-related enzymes such as chitinases and peroxidases [65, 66].

Trichoderma-mediated defense responses commonly involve induced systemic resistance [ISR] and systemic acquired resistance (SAR), which are regulated through jasmonic acid, ethylene, and salicylic acid signaling pathways [67]. For example, *T. harzianum* has been reported to induce jasmonic acid- and ethylene-mediated resistance in grapevine against *Plasmopara viticola* [68, 69]. Similarly, increased activities of defense enzymes in roots and leaves have been observed following *Trichoderma* colonization, indicating enhanced systemic resistance against pathogens [70]. The plant defence signalling pathways induced by *Trichoderma* is illustrated in figure 2.

3.5. Entomopathogenic and Nematicidal Activities

Besides suppressing plant pathogens, *Trichoderma* spp. also exhibit entomopathogenic and nematicidal properties against various agricultural pests. Entomopathogenic activity involves direct infection and colonization of insect hosts through the cuticle, followed by proliferation within the insect body and eventual death of the host [71]. Certain strains of *Trichoderma* produce secondary metabolites and enzymes with insecticidal, antifeedant, and repellent properties that reduce pest survival and feeding activity [47].

For instance, culture filtrates of *T. harzianum* containing chitinase enzymes have shown antifeedant activity against *Helicoverpa armigera* in cotton [72]. Similarly, *T. atroviride* and *T. hamatum* have demonstrated pathogenic effects against insect pests such as *Aedes aegypti* and *Spodoptera littoralis* [59, 60]. The examples of insects controlled by *Trichoderma* spp. are presented in table 2.

Trichoderma spp. are also effective against plant-parasitic nematodes [73]. Their nematicidal activity is mainly associated with the production of hydrolytic enzymes and toxic metabolites that degrade nematode eggs and inhibit larval development [74]. Studies have shown that *T. harzianum* can parasitize nematode eggs through chitinase production, significantly reducing nematode reproduction and survival [75]. In addition, non-volatile metabolites produced by *Trichoderma* spp. have been reported to cause substantial mortality in *Pratylenchus brachyurus* and *Meloidogyne incognita*, highlighting their potential as eco-friendly alternatives to chemical nematicides [55, 76].

4. Agricultural Applications of *Trichoderma*

4.1. *Trichoderma* as plant growth promoter

Trichoderma spp. produces various phytohormones ,and

compounds that helps in growth promotion [77]. The phytohormones such as gibberellic acid (GA_3) and indole-3-acetic acid (IAA) increases the expression of growth-related genes which promote root development and biomass accumulation [78]. The inoculation of *Trichoderma* spp. increase biomass and root length of *A. euteiches*-infected plants [79]. Furthermore, *Trichoderma* application enhance fruit quality such as diameter, weight, circumference and TSS [80]. Especially, growth promoting effect is very beneficial when used to use with horticulture crops with week seed vigour [81].

4.2. *Trichoderma* as biofertilizer

Trichoderma serves as a natural substitute to synthetic fertilizers due to its ability to synthesis plant growth hormones, solubilize the insoluble phosphates, and increase the uptake of micro and macro nutrients [17]. *Trichoderma* spp. solubilize several soil nutrients that are present in an inaccessible form and convert them into plant available form, which increases the O_2 and CO_2 utilization [46]. When phosphorus binds with soil and form toxic complexes in acid soil, making the phosphorus unavailable to the plants [17]. *Trichoderma* strains can produce several enzymes which solubilize insoluble phosphates in acidic soil and micronutrients [9, 77]. For instance, *T. harzianum* can increase the various nutrient availability such as solubilize various plant nutrients, such as rock phosphate, iron, copper, manganese and zinc, especially in nutrient deficient soil [82]. Similarly, Application of *Trichoderma*-based-fertilizers at the rate of 9000 kg ha⁻¹ significantly enhanced aboveground biomass in grassland system [83]. Due to the effective nutrient mobilization, roots in association with these fungi frequently are larger and more robust and consequently results in increased yield [84].

4.3. *Trichoderma* as bioremediator

Bioremediation is the process by which toxic chemicals from contaminated from soil and water are removed using biological agents, mainly microorganisms that are native to the contaminated sites [85, 86]. *Trichoderma* spp. can be used as myco-remediation of environmental contaminants of heavy metals such as lead [87]. They produce various organic acids including gluconic, fumaric and citric acid, and aids in lowering the pH of the soil, promoting the dissolution of phosphate, and micro and macronutrients like iron, magnesium which are essential for the plant metabolism. Additionally, they have extracellular enzymes within their system and catalytic reaction which promote the degradation of toxic aromatic compounds present in the soil [88]. The study conducted by Nykiel-Szymańska et al. [89] demonstrated that *Trichoderma* spp. help in the biodegradation of 80–99% of alachlor and 40–79% of metolachlor within a week.

4.4. *Trichoderma* as stress tolerant

Their colonization produces auxin and GA which enhances tolerance to other abiotic stresses through improved water and nutrient uptake due to enhanced root and shoot growth and increased chlorophyll content, eventually increasing the crop

yield [90–92]. *T. harzianum* inoculation improved drought tolerance in rice cultivation as it increases the activity of antioxidant enzymes, superoxide dismutase [SOD], catalase which eliminate the reactive oxygen species consequently preventing oxidative damage to rice due to drought [93]. Similarly, *T. asperellum* inoculation with reduced the adverse effect of drought and low temperature in tomato plant and increased the yield [94, 95]. During stress condition, it produced chlamydospores, which regulate oxygen utilization and withstand acidic condition [96]. The production of antioxidant and phenolic substances lower the leaf temperature and deal with heat stress [97].

5. Factors affecting the activities of *Trichoderma*

The efficacy of any biocontrol agent depends on the environment that they are subjected to. The full potential can only be exploited by studying the environmental conditions that favors its growth and colonization leading to proliferation [98]. Parameters such as species of *Trichoderma* and host plant and environmental conditions like temperature, pH, moisture, salinity and nutrient availability significantly affects the effectiveness of *Trichoderma* in biocontrol mechanisms [34,39,99].

5.1. Temperature

Temperature significantly affects the growth and biocontrol efficiency of *Trichoderma*. Various studies have reported the efficiency temperature range between 25-30°C as the favourable range for the maximum antagonism and the temperature less than 20°C and more than 35°C retard the growth, colonization and biocontrol properties of *Trichoderma* [33]. For instance, antagonistic activity of *Trichoderma* spp. was decreased against *Macrophomina phaseolina* when temperature was increased from 25 to 35°C [100].

5.2. Soil pH

The biocontrol property is affected by pH condition as the production of secondary metabolites is greatly influenced by the pH range and its production is higher under optimum pH [3]. Although most of species can tolerant broad pH range, it has its optimum growth under acidic conditions, at the pH ranging from 5-7 [33, 34]. Moreover, pH 6.5 was favorable for the peak growth and sporulation of *Trichoderma* along with the highest maximum inhibition of five phytopathogen in chickpea rhizosphere i.e., 63.23%, 65.85%,53.33%, 53.84%, and 48.00% against *Fusarium oxysporum* f. *ciceri*, *Sclerotium rolfsii*, *Sclerotinia sclerotiorum*, *Pythium* spp. and *Rhizoctonia bataticola*, respectively [101].

5.3. Moisture Availability

Soil and surface moisture strongly affects the sporulation, germination, secretion of metabolites and survival of *Trichoderma* [1]. Sufficient water availability is crucial for hyphal contact, coiling and penetration. *Trichoderma* can withstand the relative humidity of 20-80%, however the maximum conidial formation and survival was obtained at

70%, demonstrating the role of irrigation for maintain optimum moisture level for field application. Under moist soils maintained at 25-35°C, *Trichoderma*, completely suppressed *F. oxysporum* on chickpea over six months. On contrary, water logged and soil dry reduced its efficiency as pathogen survival is highest such condition [102]. The biocontrol efficiency increases at the moisture level between 8% against *Rhizoctonia solani* highlighting the role of optimum moisture level for disease control [103].

5.4. Soil organic matter and Nutrient Status

Organic matter improves soil physical, chemical and biological properties, thereby provide favourable micro environment conditions for *Trichoderma* growth and disease suppression [43]. Adequate organic matter and nutrient availability such as cow dung and compost manure in soil provides essential nutrient to *Trichoderma*, which facilitate its growth and survival, ultimately enhancing its activity [37]. For instance, the conjoint application of organic manure and increased nitrogen, phosphorus uptake, contributing to the maximum yield in wheat [104]. Additionally, incorporation of organic matter in soil promote the other beneficial microorganisms in soil, which synergistically enhance the efficiency of *Trichoderma* [105].

5.5. Host specificity and strain variability

Species and strain variability, and host specificity plays a significant role in the effectiveness of *Trichoderma*. *Trichoderma atroviride* was less effective than *T. harzianum* in promoting stem and canopy growth and controlling *Botrytis cinerea* in tomato plants [106]. On contrary, *T. atroviride* was found to be more effective than *T. harzianum* in controlling *Botrytis cinerea* in strawberry plants [107]. From these findings it can be inferred that the efficiency of *Trichoderma* is largely dependent on the crop species, suggesting that response to *Trichoderma* spp. is under genetic control. Therefore, it is evident to have molecular characterization of *Trichoderma* before making a general recommendation [92].

6. Challenges and Future Prospects

Despite the remarkable potential of *Trichoderma* spp. in sustainable agriculture, several challenges still limit their consistent performance and large-scale adoption under field conditions [29]. Although many studies have demonstrated promising results under laboratory and greenhouse environments, the effectiveness of *Trichoderma*-based products in open field conditions often varies due to environmental fluctuations, soil characteristics, host specificity, and interactions with native microbial communities [39, 99]. Therefore, improving the reliability and adaptability of *Trichoderma* formulations remains an important research priority.

One of the major limitations associated with *Trichoderma*-based bioinoculants is their relatively short shelf life and reduced survival under unfavorable environmental conditions such as extreme temperatures, low moisture, ultraviolet radiation, and fluctuating soil pH [33].

Environmental stress can significantly affect spore viability, rhizosphere colonization, metabolite production, and antagonistic activities, ultimately reducing field efficacy [98]. Consequently, future research should focus on developing stress-tolerant strains and improved formulations capable of maintaining stability and viability during storage and field application.

Another important challenge is strain specificity and host-dependent performance. Different *Trichoderma* species and strains exhibit varying degrees of effectiveness depending on the crop species, pathogen type, and environmental conditions [106]. For example, certain strains may perform efficiently in one crop system but show limited effectiveness in another. Advances in molecular biology, genomics, transcriptomics, and metabolomics may facilitate the identification of highly efficient strains with enhanced biocontrol and plant growth-promoting capabilities [108]. The formulation and application methods of *Trichoderma* products also require further optimization. Inconsistent field performance is often associated with inappropriate carrier materials, low shelf stability, inadequate inoculum concentration, and inefficient delivery methods [29]. Future studies should therefore focus on developing advanced formulations such as encapsulated spores, nano-formulations, and slow-release carriers to improve shelf life, colonization efficiency, and persistence in soil environments [38, 109].

Commercialization and farmer adoption of *Trichoderma*-based products remain another important concern. Limited awareness among farmers, inconsistent product quality, lack of uniform registration process, and regulatory challenges often hinder widespread use of microbial biocontrol agents, particularly in developing countries [10, 110]. Therefore, establishing quality control standards, improving farmer awareness programs, and strengthening policy support for biological alternatives are essential for successful commercialization and adoption.

Future research should also explore the genetic improvement of *Trichoderma* strains through molecular breeding and genetic engineering approaches to enhance stress tolerance, antagonistic activity, nutrient solubilization, and environmental adaptability. Furthermore, understanding the molecular interactions between *Trichoderma*, plants, pathogens, and soil microbiomes will provide valuable insights into improving its agricultural applications [77]. Emerging technologies such as omics-based approaches, artificial intelligence-assisted strain screening, and microbiome engineering may further accelerate the development of next-generation *Trichoderma*-based bioinoculants.

Although microbial consortia involving *Trichoderma*, *Pseudomonas* spp., *Bacillus* spp., and arbuscular mycorrhizal fungi are reported to have synergistic interactions in enhancing plant growth and disease suppression, can enhance plant growth and disease suppression through synergistic interactions [111, 112], antagonistic relationships among consortium members have also been reported, underscoring the significance of compatibility assessment prior to application [113]. To achieve optimum field performance, future research should emphasize on understanding these

interactions and developing optimized, crop-specific microbial consortia.

Overall, *Trichoderma* spp. hold enormous promise as multifunctional agents for sustainable agriculture. However, overcoming limitations related to environmental adaptability, formulation stability, field consistency, and commercialization will be crucial for maximizing their agricultural potential and promoting their large-scale integration into eco-friendly farming systems.

7. Conclusion

The increasing environmental and health concerns associated with the excessive use of chemical fertilizers and pesticides have accelerated the search for sustainable and eco-friendly agricultural alternatives. In this context, *Trichoderma* spp. have emerged as highly effective multifunctional microorganisms with significant potential to support sustainable crop production systems. Their versatile mechanisms, including competition, mycoparasitism, antibiosis, induction of plant defense responses, nutrient mobilization, phytohormone production, and biodegradation of toxic compounds, contribute substantially to plant health and productivity.

Beyond their role as biocontrol agents against fungal pathogens, nematodes, and insect pests, *Trichoderma* spp. also function as plant growth promoters, biofertilizers, stress alleviators, and bioremediation agents. Their ability to enhance nutrient uptake, stimulate root development, improve tolerance against abiotic stresses, and restore contaminated soils highlights their broad applicability in sustainable agriculture. Furthermore, the eco-friendly nature of *Trichoderma*-based technologies offers an effective approach to reducing dependency on synthetic agrochemicals while maintaining crop productivity and environmental safety. However, despite their considerable potential, several challenges remain regarding field consistency, environmental adaptability, shelf stability, formulation development, and strain specificity. The effectiveness of *Trichoderma* is greatly influenced by environmental conditions, host plant interactions, and application methods, emphasizing the need for precise strain selection and improved formulation technologies. Future research should therefore focus on the development of stress-tolerant and highly efficient strains, advanced delivery systems, molecular characterization, and integration with other sustainable agricultural practices.

In conclusion, *Trichoderma* spp. represent promising biological tools for achieving environmentally sustainable and economically viable agriculture. Their multifunctional roles in crop protection, plant growth promotion, nutrient management, and stress mitigation make them valuable components of integrated farming systems aimed at improving food security while preserving environmental and human health.

Author contributions

Sujata Nepal: Conceptualization, literature review, writing—original draft preparation. Surakshya Sharma: Review and

critical revision. Niraj Mahato: Review and editing. All authors read and approved the final manuscript.

Ethical approval

Not applicable.

Conflicts of Interest

The authors report no conflicts of interest.

Acknowledgment

The authors sincerely acknowledged the participants who voluntarily contributed to this review work.

Data availability statement

Data sharing is not applicable to this article as no new data were generated or analyzed in this review.

Funding

No funding received for conducting this study

References

1. Papavizas GC. *Trichoderma* and *Gliocladium*: biology, ecology, and potential for biocontrol. *Annual Review of Phytopathology*. 1985;23(1):23–54. <https://doi.org/10.1146/annurev.py.23.090185.000323>
2. Tian M, Zheng Y, Sun X, Zheng H. A research on promoting chemical fertiliser reduction for sustainable agriculture purposes: Evolutionary game analyses involving ‘government, farmers, and consumers.’ *Ecological Indicators*, 2022;144:109433. <https://doi.org/10.1016/j.ecolind.2022.109433>
3. Zañafano L, Carro-Huerga G, Rodríguez-González Á, Ramírez-Lozano D, Mayo-Prieto S, Gutiérrez H, et al. Biosolutions from Native *Trichoderma* Strains Against Grapevine Trunk Diseases. Vol. 15, *Agronomy*. 2025. p. 1901. <https://doi.org/10.3390/agronomy15081901>
4. Chou C, Castilla N, Hadi B, Tanaka T, Chiba S, Sato I. Rice blast management in Cambodian rice fields using *Trichoderma harzianum* and a resistant variety. *Crop Protection*. 2020;135:104864. <https://doi.org/10.1016/j.cropro.2019.104864>
5. Tilman D, Cassman K, Matson P, Naylor R, Polasky S. review article Agricultural sustainability and intensive production practices. *Nature*. 2002 Aug 8;418:671–7. <https://doi.org/10.1038/nature01014>
6. Ayeni O, Olagoke-Komolafe O. Environmental impact of modern agricultural practices: Strategies for reducing carbon footprint and promoting conservation. *International Journal of Management & Entrepreneurship Research*, 2024 Sep 23;6:3082–95. <https://doi.org/10.51594/ijmer.v6i9.1581>
7. Shuqin J, Fang Z. Zero Growth of Chemical Fertilizer and Pesticide Use: China’s Objectives, Progress and Challenges. *J Resour Ecol* [J]. 2018 Jan 1;9(1):50–8. <https://doi.org/10.5814/j.issn.1674-764x.2018.01.006>
8. Deising HB, Reimann S, Peil A, Weber WE. Disease Management of Rusts and Powdery Mildews BT - Agricultural Applications. In: Kempken F, editor. Berlin, Heidelberg: Springer Berlin Heidelberg; 2002. p. 243–69. https://doi.org/10.1007/978-3-662-03059-2_13
9. Joo JH, Hussein KA. Biological control and plant growth

- antagonistic properties of volatile organic compound-producing Trichoderma spp. *Frontiers in Plant Science*, 2022;13:897668. <https://doi.org/10.3389/fpls.2022.897668>
10. Gohel NM, Raghunandan BL, Patel NB, Parmar H V, Raval DB. Role of Fungal Biocontrol Agents for Sustainable Agriculture. *Fungal diversity, ecology and control management* [J]. Springer; 2022. p. 577–606. https://doi.org/10.1007/978-981-16-8877-5_28
 11. Fenibo EO, Ijoma GN, Matambo T. Biopesticides in Sustainable Agriculture: A Critical Sustainable Development Driver Governed by Green Chemistry Principles. *Frontiers in Sustainable Food Systems*, 2021;5. <https://doi.org/10.3389/fsufs.2021.619058>
 12. Abirami S, Gayathri SS, Usha C. Trichoderma as biostimulant—a plausible approach to alleviate abiotic stress for intensive production practices. *New and Future Developments in Microbial Biotechnology and Bioengineering*. Elsevier; 2022. p. 57–84. <https://doi.org/10.1016/B978-0-323-85577-8.00004-4>
 13. Mukherjee PK, Horwitz BA, Singh US, Mala Mukherjee MM, Schmoll M. Trichoderma in agriculture, industry and medicine: an overview. *CABI*. 2013;1–9. <https://doi.org/10.1079/9781780642475.0001>
 14. Pacheco-Trejo J, Aquino-Torres E, Reyes-Santamaría MI, Islas-Pelcastre M, Pérez-Ríos SR, Madariaga-Navarrete A, et al. Plant Defensive Responses Triggered by Trichoderma spp. as Tools to Face Stressful Conditions. Vol. 8, *Horticulturae*. 2022. p. 1181. <https://doi.org/10.3390/horticulturae8121181>
 15. Brotman Y, Landau U, Cuadros-Inostroza A, Takayuki T, Fernie AR, Chet I, et al. Trichoderma-plant root colonization: escaping early plant defense responses and activation of the antioxidant machinery for saline stress tolerance. *PLoS Pathogens*. 2013;9(3):e1003221. <https://doi.org/10.1371/journal.ppat.1003221>
 16. López-Bucio J, Pelagio-Flores R, Herrera-Estrella A. Trichoderma as biostimulant: exploiting the multilevel properties of a plant beneficial fungus. *Scientia Horticulturae*, (Amsterdam). 2015;196:109–23. <https://doi.org/10.1016/j.scienta.2015.08.043>
 17. Kubheka B, Ziena L. Trichoderma : A Biofertilizer and a Bio-Fungicide for Sustainable Crop Production. *Trichoderma*. 2022. <https://doi.org/10.5772/intechopen.102405>
 18. Yao X, Guo H, Zhang K, Zhao M, Ruan J, Chen J. Trichoderma and its role in biological control of plant fungal and nematode disease. *Frontiers in Microbiology*. 2023;14:1160551. <https://doi.org/10.3389/fmicb.2023.1160551>
 19. Guzmán-Guzmán P, Kumar A, de los Santos-Villalobos S, Parra-Cota FI, Orozco-Mosqueda MD, Fadji AE, et al. Trichoderma Species: Our Best Fungal Allies in the Biocontrol of Plant Diseases—A Review. Vol. 12, *Plants*. 2023. p. 432. <https://doi.org/10.3390/plants12030432>
 20. Chaverri P, Castlebury LA, Overton BE, Samuels GJ. Hypocrea/Trichoderma: species with conidiophore elongations and green conidia. *Mycologia*. 2003 Nov 1;95(6):1100–40. <https://doi.org/10.1080/15572536.2004.11833023>
 21. Cai F, Druzhinina IS. In honor of John Bissett: authoritative guidelines on molecular identification of Trichoderma. *Fungal Diversity*. 2021;107(1):1–69. <https://doi.org/10.1007/s13225-020-00464-4>
 22. Druzhinina IS, Kubicek CP, Komoń-Zelazowska M, Mulaw TB, Bissett J. The Trichoderma harzianum demon: complex speciation history resulting in coexistence of hypothetical biological species, recent agamospecies and numerous relict lineages. *BMC Evolutionary Biology*. 2010;10(1):94. <https://doi.org/10.1186/1471-2148-10-94>
 23. Zhao R, Chen K, Mao L, Zhang C. Eleven new species of Trichoderma (Hypocreaceae , Hypocreales) from China. *International journal of fungal Mycology*. 2025;16(1):180–209. <https://doi.org/10.1080/21501203.2024.2330400>
 24. Athafah A, Sabah I, Alamer A, Li B, Zhang J. A new species of Trichoderma and gliotoxin role : A new observation in enhancing biocontrol potential of T . virens against Phytophthora capsici on chili pepper. *Biological Control*. 2020;145(July 2019):104261. <https://doi.org/10.1016/j.biocontrol.2020.104261>
 25. Boukaew S, Chumkaew K, Petlamul W, Srinuanpan S, Nooprom K, Zhang Z. Biocontrol effectiveness of Trichoderma asperelloides SKRU-01 and Trichoderma asperellum NST-009 on postharvest anthracnose in chili pepper. *Food Control*. 2024;163:110490. <https://doi.org/10.1016/j.foodcont.2024.110490>
 26. Ghareeb RY, Belal EB, Khateeb NMM El, Shreef BA. Utilizing bio - synthesis of nanomaterials as biological agents for controlling soil - borne diseases in pepper plants : root - knot nematodes and root rot fungus. *BMC Plant Biology*. 2024;1–22. <https://doi.org/10.1186/s12870-024-04760-y>
 27. Pacheco JC, Poltronieri AS, Porsani M V, Zawadneak MAC, Pimentel IC. Entomopathogenic potential of fungi isolated from intertidal environments against the cabbage aphid Brevicoryne brassicae (Hemiptera: aphididae). *Biocontrol Science and Technology*. 2017 Apr 3;27(4):496–509. <https://doi.org/10.1080/09583157.2017.1315053>
 28. Rahim S, Iqbal M. Exploring Enhanced Insecticidal Activity of Mycelial Extract of Trichoderma harzianum against Diuraphis noxia and Tribolium castaneum. *Sarhad Journal of Agriculture*. 2019;35(3). <https://doi.org/10.17582/journal.sja/2019/35.3.757.762>
 29. Woo SL, Ruocco M, Vinale F, Nigro M, Marra R, Lombardi N, et al. Trichoderma-based products and their widespread use in agriculture. *The Open Mycology Journal* 2014;8(1):71–126. <https://doi.org/10.2174/1874437001408010071>
 30. Abo-Elyousr KAM, Abdel-Hafez SII, Abdel-Rahim IR. Isolation of Trichoderma and Evaluation of their Antagonistic Potential against Alternaria porri. *Journal of Phytopathology*. 2014 Sep 1;162(9):567–74. <https://doi.org/10.1111/jph.12228>
 31. Howell CR. Mechanisms employed by Trichoderma species in the biological control of plant diseases: the history and evolution of current concepts. *Plant Disease*. 2003;87(1):4–10. <https://doi.org/10.1094/PDIS.2003.87.1.4>
 32. Rey M, Delgado-Jarana J, Benitez T. Improved antifungal activity of a mutant of Trichoderma harzianum CECT 2413 which produces more extracellular proteins. *Applied Microbiology and Biotechnology*. 2001;55(5):604–8. <https://doi.org/10.1007/s002530000551>
 33. Matrood AAA, Rhouma A. Environmental factors affecting Trichoderma spp . and their biocontrol potential in post - harvest disease management. *Archives of Agriculture and Environmental Science*. 2025;10(2):285–91. <https://doi.org/10.26832/24566632.2025.1002014>
 34. Al-Ani LKT. Trichoderma from Extreme Environments: Physiology, Diversity, and Antagonistic Activity BT - Extremophiles in Eurasian Ecosystems: Ecology, Diversity, and Applications. In: Egamberdieva D, Birkeland N-K, Panosyan H, Li W-J, editors. Singapore: Springer Singapore; 2018. p. 389–403. https://doi.org/10.1007/978-981-13-0329-6_14
 35. Singh S, Singh AK, Pradhan B, Tripathi S, Kumar KS, Chand S, et al. Harnessing Trichoderma Mycoparasitism as a Tool in the Management of Soil Dwelling Plant Pathogens. *Microbial Ecology*. 2024;87(1):158. <https://doi.org/10.1007/s00248-024->

- 02472-2
36. Contreras-Cornejo HA, Macías-Rodríguez L, del-Val E, Larsen J. Ecological functions of *Trichoderma* spp. and their secondary metabolites in the rhizosphere: interactions with plants. *FEMS Microbiology Ecology*. 2016 Apr;92(4):fiw036. <https://doi.org/10.1093/femsec/fiw036>
 37. Asghar W, Craven KD, Kataoka R, Mahmood A, Asghar N, Raza T, et al. The application of *Trichoderma* spp., an old but new useful fungus, in sustainable soil health intensification: A comprehensive strategy for addressing challenges. *Plant Stress*. 2024;12:100455. <https://doi.org/10.1016/j.stress.2024.100455>
 38. Vindas-reyes E, Chac R, Rivera-m W, Box CPO, Rica C, Box CPO, et al. *Trichoderma* Production and Encapsulation Methods for Agricultural Applications. *Agriengineering*. 2024;6(3):2366–84. <https://doi.org/10.3390/agriengineering6030138>
 39. Howell CR. Mechanisms employed by *Trichoderma* species in the biological control of plant diseases: The history and evolution of current concepts. *Plant Disease*. 2003;87(1):4–10. <https://doi.org/10.1094/PDIS.2003.87.1.4>
 40. Köhl J, Kolnaar R, Ravensberg WJ. Mode of action of microbial biological control agents against plant diseases: relevance beyond efficacy. *Frontiers in Plant Science*. 2019;10:845. <https://doi.org/10.3389/fpls.2019.00845>
 41. Stracquadanio C, Quiles JM, Meca G, Cacciola SO. Antifungal Activity of Bioactive Metabolites Produced by *Trichoderma asperellum* and *Trichoderma atroviride* in Liquid Medium. Vol. 6, *Journal of Fungi*. 2020. p. 263. <https://doi.org/10.3390/jof6040263>
 42. Crowley DE. Microbial Siderophores in the Plant Rhizosphere BT - Iron Nutrition in Plants and Rhizospheric Microorganisms. Barton LL, Abadia J, editors. *Iron Nutrition in Plants and Rhizospheric Microorganisms*. Dordrecht: Springer Netherlands; 2006. p. 169–98. https://doi.org/10.1007/1-4020-4743-6_8
 43. Benítez T, Rincón A, Limón MC, Codón A. Biocontrol mechanism of *Trichoderma* strains. *International Microbiology : The Official Journal of the Spanish Society for Microbiology*. 2005 Jan 1;7:249–60.
 44. Oszust K, Cybulska J, Frac M. How Do *Trichoderma* Genus Fungi Win a Nutritional Competition Battle against Soft Fruit Pathogens? A Report on Niche Overlap Nutritional Potentiates. *International Journal of Molecular Sciences*. 2020 Jun; 21(12): 4235. <https://doi.org/10.3390/ijms21124235>
 45. Wang C, Zhuang W-Y. Carbon metabolic profiling of *Trichoderma* strains provides insight into potential ecological niches. *Mycologia*. 2020 Mar 3;112(2):213–23. <https://doi.org/10.1080/00275514.2019.1698246>
 46. Silva RN, Monteiro VN, Steindorff AS, Gomes EV, Noronha EF, Ulhoa CJ. *Trichoderma*/pathogen/plant interaction in pre-harvest food security. *Fungal Biology*. 2019;123(8):565–83. <https://doi.org/10.1016/j.funbio.2019.06.010>
 47. Poveda J. *Trichoderma* as biocontrol agent against pests: New uses for a mycoparasite. *Biological Control*. 2021;159:104634. <https://doi.org/10.1016/j.biocontrol.2021.104634>
 48. Tyśkiewicz R, Nowak A, Ozimek E, Jaroszuk-Ścisiel J. *Trichoderma*: The current status of its application in agriculture for the biocontrol of fungal phytopathogens and stimulation of plant growth. *International Journal of Molecular Sciences*. 2022; 23(4): 2329. <https://doi.org/10.3390/ijms23042329>
 49. Oyesola OL, Tonjock RK, Bello OA, Taiwo OS, Obembe OO. *Trichoderma*: a review of its mechanisms of action in plant disease control [preprint]. *Preprints.org*. 2024 May 21. <https://doi.org/10.20944/preprints202405.1378.v1>
 50. Laib DE, Benzara A, Akkal S, Bensouici C. The anti-acetylcholinesterase, insecticidal and antifungal activities of the entophytic fungus *Trichoderma* sp. isolated from *Ricinus communis* L. against *Locusta migratoria* L. and *Botrytis cinerea* Pers.: *Fr. Acta Scientifica Naturalis*. 2020;7(1):112–25. <https://doi.org/10.2478/asn-2020-0011>
 51. Macías-Rodríguez L, Guzmán-Gómez A, García-Juárez P, Contreras-Cornejo HA. *Trichoderma atroviride* promotes tomato development and alters the root exudation of carbohydrates, which stimulates fungal growth and the biocontrol of the phytopathogen *Phytophthora cinnamomi* in a tripartite interaction system. *FEMS Microbiology Ecology*. 2018 Sep 1; 94(9): fiy137. <https://doi.org/10.1093/femsec/fiy137>
 52. Coninck E, Scauflaire J, Gollier M, Liénard C, Foucart G, Manssens G, et al. *Trichoderma atroviride* as a promising biocontrol agent in seed coating for reducing *Fusarium damping-off* on maize. *Journal of Applied Microbiology*. 2020 Sep 1;129(3):637–51. <https://doi.org/10.1111/jam.14641>
 53. Zhang F, Chen C, Zhang F, Gao L, Liu J, Chen L, et al. *Trichoderma harzianum* containing 1-aminocyclopropane-1-carboxylate deaminase and chitinase improved growth and diminished adverse effect caused by *Fusarium oxysporum* in soybean. *Journal of Plant Physiology*. 2017; 210:84–94. <https://doi.org/10.1016/j.jplph.2016.10.012>
 54. Singh V, Kumar B. A review of agricultural microbial inoculants and their carriers in bioformulation. *Rhizosphere*. 2024;29:100843. <https://doi.org/10.1016/j.rhisph.2023.100843>
 55. Moo-Koh FA, Cristóbal-Alejo J, Andrés MF, Martín J, Reyes F, Tun-Suárez JM, et al. In Vitro Assessment of Organic and Residual Fractions of Nematicidal Culture Filtrates from Thirteen Tropical *Trichoderma* Strains and Metabolic Profiles of Most-Active. Vol. 8, *Journal of Fungi*. 2022. p. 82. <https://doi.org/10.3390/jof8010082>
 56. Coppola M, Cascone P, Lelio I Di, Woo SL, Lorito M, Rao R, et al. *Trichoderma atroviride* P1 Colonization of Tomato Plants Enhances Both Direct and Indirect Defense Barriers Against Insects. *Frontiers in Physiology*. 2019;Volume 10-2019. <https://doi.org/10.3389/fphys.2019.00813>
 57. Mukherjee A, Debnath P, Ghosh SK, Medda PK. Biological control of papaya aphid (*Aphis gossypii* Glover) using entomopathogenic fungi. *International Journal of Plant Research*. 2020;33(1):1–10. <https://doi.org/10.1007/s42535-019-00072-x>
 58. Islam MS, Subbiah VK, Siddiquee S. Efficacy of Entomopathogenic *Trichoderma* Isolates against Sugarcane Woolly Aphid, *Ceratovacuna lanigera* Zehntner (Hemiptera: Aphididae). Vol. 8, *Horticulturae*. 2022. p. 2. <https://doi.org/10.3390/horticulturae8010002>
 59. Banduwardena AVRC, Mendis BAN. Evaluation of mycoparasitic *Trichoderma atroviride* and entomopathogenic *Aspergillus niger* as potential bioinsecticides against the dengue vector , *Aedes aegypti*. 2025;(April):1–14. <https://doi.org/10.3389/fcimb.2025.1502579>
 60. Lana M, Simón O, Velasco P, Rodríguez VM, Caballero P, Poveda J. First study on the root endophytic fungus *Trichoderma hamatum* as an entomopathogen: Development of a fungal bioinsecticide against cotton leafworm (*Spodoptera littoralis*). *Microbiological Research*. 2023 May;270:127334. <https://doi.org/10.1016/j.micres.2023.127334>
 61. Kushiyev R, Tuncer C, Erper I, Özer G. The utility of *Trichoderma* spp . isolates to control of *Xylosandrus germanus* Blandford (Coleoptera : Curculionidae : Scolytinae). *Journal of Plant Disease and Protection*. 2020; (0123456789). <https://doi.org/10.1007/s41348-020-00375-1>

62. Elsherbiny EA, Amin BH, Aleem B, Kingsley KL, Bennett JW. Trichoderma Volatile Organic Compounds as a Biofumigation Tool against Late Blight Pathogen *Phytophthora infestans* in Postharvest Potato Tubers. *Journal of Agricultural and Food Chemistry*. 2020 Aug 5; 68 (31): 8163–71. <https://doi.org/10.1021/acs.jafc.0c03150>
63. Al-Hazmi AS, TariqJaveed M. Effects of different inoculum densities of *Trichoderma harzianum* and *Trichoderma viride* against *Meloidogyne javanica* on tomato. *Saudi Journal of Biological Sciences*. 2016; 23(2): 288–92. <https://doi.org/10.1016/j.sjbs.2015.04.007>
64. Pocurull M, Fullana AM, Ferro M, Valero P, Escudero N, Saus E, et al. Commercial formulates of *Trichoderma* induce systemic plant resistance to *Meloidogyne incognita* in tomato and the effect is additive to that of the Mi-1.2 resistance gene. *Frontiers in Microbiology*. 2020; 10:3042. <https://doi.org/10.3389/fmicb.2019.03042>
65. Aljbory Z, Chen M-S. Indirect plant defense against insect herbivores: a review. *Insect Science*. 2018 Feb;25(1):2–23. <https://doi.org/10.1111/1744-7917.12436>
66. Juan CA, Pérez de la Lastra JM, Plou FJ, Pérez-Lebeña E. The Chemistry of Reactive Oxygen Species (ROS) Revisited: Outlining Their Role in Biological Macromolecules (DNA, Lipids and Proteins) and Induced Pathologies. Vol. 22, *International Journal of Molecular Sciences*. 2021. p. 4642. <https://doi.org/10.3390/ijms22094642>
67. Saldajeno MGB, Naznin HA, Elsharkawy MM, Shimizu M, Hyakumachi M. Chapter 35 - Enhanced Resistance of Plants to Disease Using *Trichoderma* spp. Gupta VK, Schmoll M, Herrera-Estrella A, Upadhyay RS, Druzhinina I, Tuohy MGBT-B and B of T, editors. Amsterdam: Elsevier; 2014. p. 477–93. <https://doi.org/10.1016/B978-0-444-59576-8.00035-7>
68. Perazzolli M, Dagostin S, Ferrari A, Elad Y, Pertot I. Induction of systemic resistance against *Plasmopara viticola* in grapevine by *Trichoderma harzianum* T39 and benzothiadiazole. *Biological Control*. 2008; 47(2):228–34. <https://doi.org/10.1016/j.biocontrol.2008.08.008>
69. Perazzolli M, Roatti B, Bozza E, Pertot I. *Trichoderma harzianum* T39 induces resistance against downy mildew by priming for defense without costs for grapevine. *Biological Control*. 2011;58(1):74–82. <https://doi.org/10.1016/j.biocontrol.2011.04.006>
70. Yedidia I, Benhamou N, Chet I. Induction of defense responses in cucumber plants (*Cucumis sativus* L.) by the Biocontrol agent *Trichoderma harzianum*. *Applied and Environmental Microbiology*. 1999;65(3):1061–70. <https://doi.org/10.1128/aem.65.3.1061-1070.1999>
71. Khaleil M, El-Mougith A, Hashem H, Lokma N. Biocontrol potential of entomopathogenic fungus, *Trichoderma hamatum* against the cotton aphid, *Aphis Gossypii*. *Journal of Environmental Science, Toxicology and Food Technology*. 2016;10(5):11–20. <https://doi.org/10.9790/2402-105021120>
72. Binod P, Sukumaran RK, Shirke S V, Rajput JC, Pandey A. Evaluation of fungal culture filtrate containing chitinase as a biocontrol agent against *Helicoverpa armigera*. *Journal of Applied Microbiology*. 2007 Nov 1;103(5):1845–52. <https://doi.org/10.1111/j.1365-2672.2007.03428.x>
73. Ibrahim D, Elderiny M, Ansari R, Rizvi R, Sumbul A, Mahmood I. Role of *Trichoderma* spp. in the Management of Plant-Parasitic Nematodes Infesting Important Crops. *Management of Phytonematodes: Recent Advances and Future Challenges*. 2020. p. 259–78. https://doi.org/10.1007/978-981-15-4087-5_11
74. Ding H, Li X, Wang S, Yang Y, Chen X, Chen C, et al. *Trichoderma harzianum* for the control of agricultural pests: Potential, progress, applications and future prospects. *Revista Argentina de Microbiología*. 2026;58(1):101–18. <https://doi.org/10.1016/j.ram.2025.09.004>
75. Oliveira CM, Oshiquiri LH, Almeida NO, Steindorf AS, da Rocha MR, Georg RC, et al. *Trichoderma harzianum* transcriptome in response to the nematode *Pratylenchus brachyurus*. *Biological Control*. 2023;183:105245. <https://doi.org/10.1016/j.biocontrol.2023.105245>
76. Oliveira C, Almeida N, De Carvalho Barros Côrtes MV, Lobo Junior M, Rocha M, Ulhoa C. Biological control of *Pratylenchus brachyurus* with isolates of *Trichoderma* spp. on soybean. *Biological Control*. 2020 Sep 1;152:104425. <https://doi.org/10.1016/j.biocontrol.2020.104425>
77. Macías-Rodríguez L, Contreras-Cornejo HA, Adame-Garnica SG, Del-Val E, Larsen J. The interactions of *Trichoderma* at multiple trophic levels: inter-kingdom communication. *Microbiological Research*. 2020;240:126552. <https://doi.org/10.1016/j.micres.2020.126552>
78. Contreras-Cornejo HA, Macías-Rodríguez L, Cortés-Penagos C, López-Bucio J. *Trichoderma virens*, a Plant Beneficial Fungus, Enhances Biomass Production and Promotes Lateral Root Growth through an Auxin-Dependent Mechanism in *Arabidopsis*. *Plant Physiology*. 2009 Mar 1;149(3):1579–92. <https://doi.org/10.1104/pp.108.130369>
79. Bazghaleh N, Prashar P, Woo S, Vandenberg A. Effects of lentil genotype on the colonization of beneficial *Trichoderma* Species and biocontrol of *Aphanomyces* Root Rot. Vol. 8, *Microorganisms*. 2020. p. 1290. <https://doi.org/10.3390/microorganisms8091290>
80. Nuangmek W, Aiduang W, Kumla J, Lumyong S, Suwannarach N. Evaluation of a newly identified endophytic fungus, *Trichoderma phayaense* for plant growth promotion and biological control of gummy stem blight and wilt of muskmelon. *Frontiers in Microbiology*. 2021;12:634772. <https://doi.org/10.3389/fmicb.2021.634772>
81. Björkman T, Blanchard LM, Harman GE. Growth enhancement of shrunken-2 (sh2) sweet corn by *Trichoderma harzianum* T295-22: Effect of environmental stress. Vol. 123, *Journal of the American Society for Horticultural Science*. 1998. p. 35–40. <https://doi.org/10.21273/jashs.123.1.35>
82. Contreras A. Ecological functions of *Trichoderma* spp. and their secondary metabolites in the rhizosphere: Interactions with plants. *FEMS microbiology ecology* 2016; 92(4). <https://doi.org/10.1093/femsec/fiw036>
83. Zhang F, Huo Y, Cobb AB, Luo G, Zhou J, Yang G, et al. *Trichoderma* biofertilizer links to altered soil chemistry, altered microbial communities, and improved grassland biomass. *Frontiers in Microbiology*. 2018;9(APR):1–11. <https://doi.org/10.3389/fmicb.2018.00848>
84. Harman GE. Myths and Dogmas of Biocontrol Changes in Perceptions Derived from Research on *Trichoderma harzianum* T-22. *Plant Disease*. 2000 Apr 1;84(4):377–93. <https://doi.org/10.1094/PDIS.2000.84.4.377>
85. Strong J, Burgess J. Treatment Methods for Wine-Related and Distillery Wastewaters: A Review. *Bioremediation Journal*. 2008 May 22;12(1):70–87. <https://doi.org/10.1080/10889860802060063>
86. Shakshi, Singh S. , Haritash A. Environmental Biotechnology for control of environmental pollution. *International Journal of Advanced Research* 2018;6(11):816–9. <https://doi.org/10.21474/IJAR01/8064>
87. Bandurska K, Krupa P, Berdowska A, Jatulewicz I, Zawierucha I. Mycoremediation of soil contaminated with cadmium and lead by *Trichoderma* sp. *Ecological Chemistry and Engineering*. 2021;28(2):277–86.

- <https://doi.org/10.2478/eces-2021-0020>
88. Kumar M, Fatehpuria PK, Ahmad SK, Jamil A, Dhakar N. Application of *Trichoderma* spp. Restoration in Soil Health. *International Journal of Current Microbiology and Applied Sciences*. 2020;9(2):3051–8. <https://doi.org/10.20546/ijcmas.2020.902.351>
 89. Nykiel-Szymańska J, Bernat P, Slaba M. Biotransformation and detoxification of chloroacetanilide herbicides by *Trichoderma* spp. with plant growth-promoting activities. *Pesticide Biochemistry and Physiology*. 2020;163:216–26. <https://doi.org/10.1016/j.pestbp.2019.11.018>
 90. Bae H, Sicher RC, Kim MS, Kim SH, Strem MD, Melnick RL, et al. The beneficial endophyte *Trichoderma hamatum* isolate DIS 219b promotes growth and delays the onset of the drought response in *Theobroma cacao*. *Journal of Experimental Botany*. 2009;60(11):3279–95. <https://dx.doi.org/10.1093/jxb/erp165>
 91. Khan MIR, Fatma M, Per TS, Anjum NA, Khan NA. Salicylic acid-induced abiotic stress tolerance and underlying mechanisms in plants. *Frontiers in Plant Science*. 2015;6:462. <https://doi.org/10.3390/pathogens10080991>
 92. Illescas M, Pedrero-Méndez A, Pitorini-Bovolini M, Hermosa R, Monte E. Phytohormone Production Profiles in *Trichoderma* Species and Their Relationship to Wheat Plant Responses to Water Stress. Vol. 10, *Pathogens*. 2021. p. 991. <https://doi.org/10.3390/pathogens10080991>
 93. Gusain DY, Singh U, Sharma A. Enhance activity of stress related enzymes in rice (*Oryza sativa* L.) induced by plant growth promoting fungi under drought stress. *African Journal of Agricultural Research*. 2014 May 19;9(10):1430–4. <https://doi.org/10.5897/AJAR2014.8575>
 94. Cornejo-Ríos K, Osorno-Suárez MD, Hernández-León S, Reyes-Santamaría MI, Juárez-Díaz JA, Pérez-España VH, et al. Impact of *Trichoderma asperellum* on chilling and drought stress in Tomato (*Solanum lycopersicum*). Vol. 7, *Horticulturae*. 2021. p. 385. <https://doi.org/10.3390/horticulturae7100385>
 95. Ghorbanpour A, Salimi A, Ghanbary MAT, Pirdashti H, Dehestani A. The effect of *Trichoderma harzianum* in mitigating low temperature stress in tomato (*Solanum lycopersicum* L.) plants. *Scientia Horticulturae*. 2018;230(7):134–41. <https://doi.org/10.1016/j.scienta.2017.11.028>
 96. Houlton BZ, Almaraz M, Aneja V, Austin AT, Bai E, Cassman KG, et al. A world of co-benefits: Solving the global nitrogen challenge. *Earth's Future*. 2019;7:1–8. <https://doi.org/10.1029/2019ef001222>
 97. Banjade D, Khanal D, Regmi P, Shrestha A, Banjade N. Mitigating heat stress in tomato by synergetic effect of *Trichoderma* and organic manures. *Journal of Agriculture and Natural Resources*. 2024;7(1):50–61. <https://doi.org/10.3126/janr.v7i1.73124>
 98. Bhai RS, Raj S, Kumar A, Calicut MPO. Influence of soil pH and moisture on the biocontrol potential of *Trichoderma harzianum* on *Phytophthora capsici* – black pepper system. *Journal of Biological Control*. 2010;24(2):153–7. <https://doi.org/10.18311/jbc/2010/3600>
 99. Zin NA, Badaluddin NA. Biological functions of *Trichoderma* spp. for agriculture applications. *Annals of Agricultural Sciences*. 2020;65(2):168–78. <https://doi.org/10.1016/j.aos.2020.09.003>
 100. Malathi P, Sabitha Doraisamy SD. Effect of temperature on growth and antagonistic activity of *Trichoderma* spp. against *Macrophomina phaseolina*. *Journal of Biological Control*. 2003;17(2):153–159. <https://doi.org/10.18311/jbc/2003/3973>
 101. Maurya MK, Srivastava M. Effect of various pH levels on the growth and sporulation of *Trichoderma viride* isolates and assessing their antagonistic activity against soil-borne pathogens. *Journal of Pure and Applied Microbiology*. 2024;18(November):2516–27. <https://doi.org/10.22207/JPAM.18.4.23>
 102. Inam-Ul-Haq M, Javed N, Khan MA, Jaskani MJ, Khan MM, Khan HU, et al. Role of temperature, moisture and *Trichoderma* species on the survival of *Fusarium oxysporum* ciceri in the rainfed areas of Pakistan. *Pakistan Journal of Botany*. 2009;41(4):1965–74. [https://mail.pakbs.org/pjbot/PDFs/41\(4\)/PJB41\(4\)1965.pdf](https://mail.pakbs.org/pjbot/PDFs/41(4)/PJB41(4)1965.pdf)
 103. Nadeem A, Hussain S, Akbar A, Ahmad Z, Tariq M, Huzaifa M. Effect of different levels of soil moisture and *Trichoderma viride* in controlling black scurf of potato. *Sarhad Journal of Agriculture*. 1991;38(1):40–5. <https://doi.org/10.17582/journal.sja/2022/38.1.40.45>
 104. Ren L, Lv J, Zhang F, Dou B, Li L, Wang Y, et al. Integrated fertilization with organic manure and *Trichoderma* enhances wheat productivity and soil nutrient availability. *Frontiers in Plant Science*. 2025;16(October):1–14. <https://doi.org/10.3389/fpls.2025.1687216>
 105. Hoitink HAJ, Boehm M. Biocontrol Within the Context of Soil Microbial Communities: A Substrate-Dependent Phenomenon. *Annual Review of Phytopathology*. 1999 Feb 1;37:427–46. <https://doi.org/10.1146/annurev.phyto.37.1.427>
 106. Tucci M, Ruocco M, De Palma L, De Lorito M. The beneficial effect of *Trichoderma* spp. on tomato is modulated by the plant genotype. *Molecular Plant Pathology*. 2011;12(4):341–54. <https://doi.org/10.1111/j.1364-3703.2010.00674.x>
 107. Ahmed M, Ali A, Shaheen I. Efficacy of some biological control treatments on grey mold disease (*Botrytis cinerea*) of strawberry fruits. *Journal of Plant Protection and Pathology*. 2023 Nov 12;14(11):345–51. <https://doi.org/10.21608/jppp.2023.242510.1187>
 108. Druzhinina IS, Seidl-Seiboth V, Herrera-Estrella A, Horwitz BA, Kenerley CM, Monte E, et al. *Trichoderma*: the genomics of opportunistic success. *Nature Reviews Microbiology*. 2011;9(10):749–59. <https://doi.org/10.1038/nrmicro2637>
 109. Brondi MG, Florencio C, Vasconcellos VM, Ribeiro C, Farinas CS. Enhancing the Shelf Life and Stress Tolerance of the Biocontrol Agent *Trichoderma harzianum* by Encapsulation in Green Matrices of Nanocellulose and Carboxymethyl Cellulose. *ACS Agriculture Science and Technology*. 2025 Jun 16;5(6):1178–88. <https://doi.org/10.1021/acsagscitech.5c00189>
 110. Kubiak A, Wolna-Maruwka A, Niewiadomska A, Pilarska AA. The Problem of Weed Infestation of Agricultural Plantations vs. the Assumptions of the European Biodiversity Strategy. Vol. 12, *Agronomy*. 2022. p. 1808. <https://doi.org/10.3390/agronomy12081808>
 111. Poveda J, Eugui D. Combined use of *Trichoderma* and beneficial bacteria (mainly *Bacillus* and *Pseudomonas*): Development of microbial synergistic bio-inoculants in sustainable agriculture. *Biological Control*. 2022;176:105100. <https://doi.org/10.1016/j.biocontrol.2022.105100>
 112. Santoyo G, Orozco-Mosqueda M del C, Afridi MS, Mitra D, Valencia-Cantero E, Macías-Rodríguez L. *Trichoderma* and *Bacillus* multifunctional allies for plant growth and health in saline soils: recent advances and future challenges. *Frontiers in Microbiology*. 2024;Volume 15-2024. <https://doi.org/10.3389/fmicb.2024.1423980>
 113. Dugassa A, Alemu T, Woldehawariat Y. In-vitro compatibility assay of indigenous *Trichoderma* and *Pseudomonas* species and their antagonistic activities against black root rot disease

(*Fusarium solani*) of faba bean (*Vicia faba* L.). BMC Microbiology. 2021;21(115):1–11.
<https://doi.org/10.1186/s12866-021-02181-7>



Copyright © 2026 by the author[s].

Published by Science Research Publishers.

This article is an open-access article distributed under the terms and conditions of the Creative Commons Attribution-NonCommercial-NoDerivatives [CC BY-NC-ND 4.0] License Visit <https://creativecommons.org/licenses/by-nc-nd/4.0/>

How to cite this article: Nepal, S., Sharma, S., & Mahato, N. (2026). Multifaceted Roles of *Trichoderma* in Sustainable Agriculture: Mechanisms, Applications, and Future Perspectives. *Journal of Soil, Plant and Environment*, 5(2), 1–13.